BCL-2 FAMILY OF PROTEINS AND CELL CYCLE REGULATORY GENES PLAY A ROLE IN THE REGULATION OF APOPTOSIS INDUCED BY LITHIUM AND CALYCULIN-A IN HL-60 CELLS

By

TSHABALALA NKHENSANI CECILIA

A thesis submitted in fulfillment of the requirements for the degree of

Master of Science

In the Department of Biochemistry, Microbiology and Biotechnology, Faculty of
Sciences and Agriculture, University of Limpopo (Turfloop Campus)

Private Bag x1106, Sovenga, 0727

South Africa

April 2007

Supervisor: Prof. LJ Mampuru

DECLARATION

I declare that the dissertation hereby submitted to the University of Limpopo for the degree Master of Science has not previously been submitted by me for a degree at this or any other University, that this is my own work in design and in execution, and that all materials contained therein have been duly acknowledged.

Signed:	 	
Date:		

DEDICATION

This thesis is dedicated to my mother Matlhamu Sellina Ngobeni and my late father Tsakani Donald Tshabalala *etlela hi kurhula nkovana*.

ACKNOWLEDGEMENTS

The bible says "the beginning of knowledge is the fear of the Lord". Lord I thank You, for the steps of righteous man are ordered by God. First and foremost I want to appreciate the following people for their outstanding support and contribution to my study:

- Let me take this opportunity to appreciate the good leadership skills of Prof LJ Mampuru. Your support and dedication in my research can not go unnoticed. God bless you and your family.
- I thank National Research Foundation (NRF) and the University of Limpopo Research and Admnistration for their financial assistance throughout these years.
- The staff of the Department of Biochemistry, Microbiology and Biotechnology for their support and advices and I believe that God will bless you.
- Alma Moller (Centre for Microscopy and Microanalysis) for assisting with the darkroom.
- I would like to express my warm gratitude to all my friends who were there when I needed them: Winnie, Mabasa Given and Dr Richard Hasani Chauke *xa humba mbhurhi ya mhlengwe*. How can I forget Sinah my friend? When I was hopeless you were there to give me strength and you listen to me when I cried, friend I thank you. I will never forget you.
- Last but not least to my family who were there for me through it all. I cannot mention your efforts because my words cannot begin to do justice to your hardwork, great and small. Mhani kuhava munhu wo fana na n'wina ka leyi misava. Xinyori uri nkhensa mutswari waha hanya. Mhani ntsongo ndza nkhensa ka leswi minga swi endla malembe hinkwawo ku sukela loko ndzi nghena xikolo.

Sesi Maria, Aretha, Ndzali, Tshikani na Xongi n'wananga inkomu ka hinkwaswo leswi minga swi endla.

"Ndzi nkhensile"

TABLE OF CONTENTS

I	PAGE
Declaration	ii
Dedication	iii
Acknowledgements	iv
Table of contents	vi
List of figures	ix
List of tables	xi
List of abbreviations	xii
Abstract	xvi
CHAPTER 1: INTRODUCTION	
1.1. Lithium	- 1
1.1.1. Mechanisms of lithium action	2
1.2. Calyculin-A	- 4
1.3. Mechanisms of cell death	- 5
1.3.1. Differences between necrosis and apoptosis	- 5
1.4. Mechanisms of apoptosis	- 6
1.4.1. Intrinsic or mitochondrial pathway	6
1.4.2. Extrinsic pathway (Death-receptor pathway)	7
1.5. Regulation of apoptosis	9
1.5.1. Bcl-2 family of proteins	9
1.5.2. Bcl-2 protein	10
1.5.3. Bax protein	12
1.5.4. Bad protein	13
1.5.5. p53 protein	14
1.6. Cell cycle (How it is related to apoptosis)	15
1.6.1. Regulation of the cell cycle	16
1.6.2. The role of p34 ^{cdc2} during cell cycle	18
1.7. Reversible phosphorylation of proteins	- 19

1.7.1. Okadaic acid	20
1.8. Protein phosphatases	21
1.8.1. Type-2A protein phosphatase (PP2A)	22
1.8.2. The role of protein phosphatases in the regulation of cell cycle and apoptosis-	24
1.8.2.1. Type-1 protein phosphatase (PP1)	24
1.8.2.2. PP2A as a positive component of cellular growth control	25
1.8.3. PP2A as a regulator of Bcl-2 family proteins and caspases	26
1.8.3.1. Inactivation of anti-apoptotic Bcl-2 protein	26
1.8.3.2. Activation of pro-apoptotic Bad protein	26
1.8.3.3. PP2A as a substrate of caspases	27
1.8.3.4. PP2A as a potential tumour suppressor or tumour promoter	27
1.9. Rationale	28
1.10. Aims and objectives	29
CHAPTER 2: MATERIALS AND METHODS	
2.1. Chemicals	30
2.2. Equipment	31
2.3. Cell culture, growth conditions and drug treatment	31
2.4. Total RNA extraction	32
2.4.1. RT-PCR of apoptotic and cell cycle regulatory genes	32
2.5. Western blotting	
2.5. Western blotting	33
-	33 33
2.5.1. Protein extraction	
2.5.1. Protein extraction	33
2.5.1. Protein extraction	33 34
2.5.1. Protein extraction	33 34
2.5.1. Protein extraction	33 34 34

cells	39
3.4. The effect of the combination of lithium and CL-A on the viability of HL-60	
cells	- 39
3.5. RT-PCR analyses of apoptotic and cell cycle regulatory genes in HL-60 cells	
after lithium treatment	- 42
3.6. RT-PCR analyses of apoptotic and cell cycle regulatory genes in HL-60 cells	
after CL-A treatment	- 44
3.7. RT-PCR analyses of apoptotic genes in HL-60 cells after treatment	
with the combination of both lithium and CL-A	40
3.8. RT-PCR analyses of cell cycle regulatory genes in HL-60 cells after lithium	
treatment	- 48
3.9. RT-PCR analyses of cell cycle regulatory genes in HL-60 cells after	
CL-A treatment	48
3.10. RT-PCR analyses of cell cycle regulatory genes in HL-60 cells	
after treatment with the combination of lithium and CL-A	52
3.11. Western blot analyses of apoptotic and cell cycle regulatory proteins in HL-6	0
cells after lithium treatment	- 52
3.12. Western blot analyses of apoptotic and cell cycle regulatory proteins	
after CL-A treatment	- 55
3.13. Western blot analyses of apoptotic and cell cycle regulatory proteins	
after combination of lithium and CL-A treatment	- 55
CHAPTER 4: DISCUSSION	59
CHAPTER 5: REFERENCES	66

LIST OF FIGURES

		TT	n		
н	(Ш	K	H۷	•

1.1. Calyculin-A structure	5
1.2. Mechanisms of cell death	
1.3. Major apoptotic pathways in mammalian cells: the intrinsic and extrinsic	
pathways	8
1.4. The balance between pro- and anti-apoptotic proteins determine the cell	
cycle decision	11
1.5. Schematic representation of cyclins/Cdk protein complexes and the	
cell cycle	. 17
1.6. PP2A holoenzyme showing the catalytic C, regulatory A and targeting	
B subunits	- 23
3.1. Cell proliferation (A) and viability (B) of HL-60 cells after treatment	
with various concentrations of lithium	36
3.2. Cell proliferation (A) and viability (B) of HL-60 cells after treatment	
with various concentrations of CL-A	38
3.3. Cell proliferation of HL-60 cells after treatment with various	
concentrations of the combination of lithium and CL-A	40
3.4. Cell viability of HL-60 cells after treatment with various concentrations of	
the combination of lithium and CL-A	41
3.5A. Total RNA isolated from HL-60 cells treated with different	
concentrations of lithium	43
3.5B. RT-PCR analyses of bax, bcl-2, cyclin-B1 and cdc2 in HL-60 cells	
after treatment with various concentrations of lithium	43
3.6A. Total RNA isolated from HL-60 cells treated with various concentrations of	
CL-A	45
3.6B. RT-PCR analyses of bax, bcl-2, cyclin-B1 and cdc2 of HL-60 cells	
after treatment with various concentrations of CL-A	45
3.7A. Total RNA isolated from HL-60 cells treated with various concentrations of	
lithium and CL-A	- 47

3.7B. RT-PCR analyses of <i>bax</i> and <i>bcl-2</i> of HL-60 cells after treatment	
with increasing concentrations of lithium and CL-A	4
3.8A. Total RNA isolated from HL-60 cells treated with various concentrations of	
lithium	49
3.8B. RT-PCR analyses of cyclin-B1 and cdc2 of HL-60 cells after treatment	
with various concentrations of lithium	49
3.9A. Total RNA isolated from HL-60 cells treated with CL-A	5(
3.9B. RT-PCR analyses of cyclin-B1 and cdc2 of HL-60 cells after treatment with	
various concentrations of CL-A	5
3.10A. Total RNA isolated from HL-60 cells treated with various concentrations	
of the combination lithium and CL-A	5
3.10B. RT-PCR analyses of cyclin-B1 and cdc2 of HL-60 cells after treatment with	
various concentrations of lithium and CL-A	5
3.11A. A Coomassie blue stained SDS-PAGE gel of the total protein from HL-60 cells	3
treated with various concentrations of lithium	5.
3.11B. Effect of lithium on the expression levels of Bcl-2 and Cdc2 proteins in	
HL-60 cells	5.
3.12A. A Coomassie blue stained SDS-PAGE gel of the total protein from HL-60 cells	3
treated with various concentrations of CL-A	5
3.12B. Effect of CL-A on the expression levels of Bcl-2 and Cdc2 proteins in	
HL-60 cells treated with various concentrations of CL-A	5
3.13A. A Coomassie blue stained SDS-PAGE gel of the total protein isolated from	
HL-60 cells treated with various concentrations of the combination of both	
lithium and CL-A	58
3.13B. Effect of combination of both lithium and CL-A on the expression levels of	
Bax, Bcl-2 and Cdc2 proteins in HL-60 cells	58

LIST OF TABLES

TA	BLE:

2.1. Biochemicals, media, cell culture and assay kits used in the study	30
2.2. Gene specific primers used for RT-PCR	33

LIST OF ABBREVIATIONS

•	Akt	-	Protein kinase B (PKB)
•	Apaf-1	-	Apoptotic protease activating factor-1
•	ATCC	-	American type culture collection
•	Bad	-	Bcl-2-X _L /Bcl-2 associated death promoter
•	Bax	-	Bcl-2 associated protein-X
•	BCA	-	Bicinchoninic acid
•	Bcl-2	-	B-cell leukaemia-2
•	$Bcl-X_L$	-	Anti-apoptotic Bcl-2 family member
•	Bcl-X _S	-	Anti-apoptotic Bcl-2 family member
•	BH1-3	-	Bcl-2 homologue regions 1, 2 and 3
•	Bid	-	BH3-interacting domain death agonist
•	Bik	-	Bcl-2 interacting killer
•	CAPS	-	3-[cyclohexylamino] -1- propanesulphonic acid
•	Cdc2	-	Cell division cycle-2 protein
•	CDKs	-	Cyclin-dependent kinases
•	cDNA	-	Complementary DNA
•	CL-A	-	Calyculin-A
•	CNS	-	Central nervous system
•	CO_2	-	Carbon dioxide
•	-COOH	-	Carboxyl group
•	°C	-	Degrees centigrade (Celsius)
•	DAG	-	Diacylglycerol
•	DISC	-	Death-inducing signaling complex
•	DNA	-	Deoxyribose nucleic acid
•	DNase	-	Deoxyribonuclease
•	EDTA	-	Ethylenediaminetetraacetic acid
•	dATP	-	Deoxyadenosine triphosphate
•	dNTPs	-	Deoxyribonucleotide triphosphates

•	FADD	-	Fas-associated death domain
•	FasL	-	Fas ligand
•	FBS	-	Foetal bovine serum
•	FLICE	-	FADD-like interleukin-1-converting enzyme
•	G0-phase	-	Gap-0 (zero) phase of the eukaryotic cell division
			cycle
•	G1-phase	-	Gap-1 phase of the eukaryotic cell division cycle
•	G2-phase	-	Gap-2 phase of the eukaryotic cell division cycle
•	GSK-3	-	Glycogen synthase kinase-3
•	h	-	Hour
•	H1	-	Histone-1
•	H_2O	-	Dihydrogen monoxide (Water)
•	HL-60	-	Human promyelocytic leukaemia cells
•	HRP	-	Horseradish peroxidase
•	I-(1-2)	-	Inhibitor (heat-stable inhibitor protein)
•	IC_{50}	-	Half maximal inhibitory concentration
•	IgG	-	Immunoglobulin G
•	IP	-	Inositol monophosphate
•	IP3	-	Inositol 1,4,5-triphosphate
•	K^{+}	-	Potassium ion
•	kDA	-	Kilodalton
•	K_{i}	-	Concentration of inhibition which doubles the
			observed K_m of the enzyme
•	\mathbf{K}_{m}	-	Michaelis constant
•	Li ⁺	-	Lithium ion
•	LiCl	-	Lithium chloride
•	mA	-	Milliamperes
•	MDM2	-	Murine double minute 2
•	ml	-	Millilitres
•	min	-	Minute

•	mM	-	Millimolar
•	MOPS	-	3-(N-morpholino) propanesulfonic acid
•	MPF	-	Mitotic phase promoting factor/ Maturation
			promoting factor
•	M-phase	-	Mitotic phase
•	mRNA	-	messenger RNA
•	MuLV	-	Murine leukaemia virus
•	Na^+	-	Sodium ion
•	NaCl	-	Sodium chloride
•	Nek 2	-	NIMA-related protein kinase 2
•	NIMA	-	Never in mitosis gene A
•	nM	-	Nanomolar
•	NP-40	-	Non-ionic detergent P-40
•	PBS	-	Phosphate-buffered saline
•	PCR	-	Polymerase chain reaction
•	PI	-	Polyphosphoinositol
•	PIP2	-	Phosphatidylinositol bisphosphate
•	PKB	-	Protein kinase B
•	PLC	-	Phospholipase C
•	PMSF	-	Phenylmethylsulfonyl fluoride
•	PP1	-	Protein phosphatase type-1
•	PP2A	-	Protein phosphatase type-2A
•	PP2B	-	Protein phosphatase type-2B
•	PP2C	-	Protein phosphatase type-2C
•	PPM	-	Mg ²⁺ -dependent protein phosphatase
•	PPP	-	Phosphoprotein phosphatase
•	pRB	-	Retinoblastoma protein
•	PSN	-	Penicillin/streptomycin/neomycin cocktail
•	PTP	-	Protein tyrosine phosphatase
•	R	-	Restriction point (cell cycle)

• RPMI - Roswell Park Memorial Institute

• RNA - Ribose nucleic acid

• RNase - Ribonuclease

• rpm - Revolutions per minute

• rRNA - Ribosomal RNA

• RT-PCR - Reverse transcription-polymerase chain reaction

• SDS - Sodium dodecyl sulphate

• SDS-PAGE - Sodium dodecylsulphate-polyacrylamide gel

electrophoresis

• sec - Second

• S-phase - DNA synthesis phase

• Ser - Serine

• TBS - Tris-buffered saline

• TEMED - Tetramethylethylenediamine

• Thr - Threonine

• TNF - Tumour necrosis factor

• TRIS - Trishydroxymethylaminomethane

• Tyr - Tyrosine

• UV - Ultraviolet

• µl - Microliter

ABSTRACT

The biochemical mechanism of apoptosis induced by lithium remains unclear, although there is evidence suggesting the involvement of Bax and Bcl-2. Bcl-2 family of proteins play a critical role in the regulation of apoptosis in various tumour cell lines. This pathway may be altered in cancer cells. We have used calyculin-A (CL-A), an inhibitor of protein phosphatase 2A (PP2A), to investigate the mechanism by which lithium induces apoptosis in HL-60 cells. Previous studies in our laboratory established that lithium induces apoptosis of HL-60 cells at 10 mM and above; while CL-A induces apoptosis at 1 nM and above. The observed apoptotic effects were additive. These observations led to the hypothesis that lithium and CL-A exert their biological effects by acting on a similar target. It was, therefore, the aim of this study to establish whether lithium would also exert similar inhibitory effects on the apoptotic and cell cycle regulatory genes. We further aimed at delineating the effects of both lithium and CL-A on the expression profiles of apoptotic and cell cycle regulatory genes. In this study, HL-60 cells were treated with lithium, CL-A and the combination of both. This was followed by the assessment of cell proliferation and viability at specific time points, using Coulter Counter and trypan blue dye exclusion assay, respectively. Concentrations of lithium at 10 mM and 20 mM were found to inhibit cell proliferation and exerted modest effects on cell viability in a time- and dose-dependent manner. Likewise, CL-A inhibited cell proliferation and viability in a time- and dose-dependent fashion. The combination of lithium and CL-A showed additive inhibitory effects on the growth of HL-60 cells.

Further, semi-quantitative RT-PCR analyses of apoptotic (*bax* and *bcl-2*) and cell cycle regulatory genes (*cdc2* and *cyclin-B1*) were determined. Our data revealed an underexpression of *bcl-2* mRNA and an up-regulation of *bax* mRNA in HL-60 cells treated with lithium, CL-A and the combination of both. In addition, the expression levels of *cdc2* mRNA remained constant, while *cyclin-B1* mRNA expression levels were up-regulated after 24 h in HL-60 cells that were treated with cytotoxic concentrations of lithium and CL-A alone. Furthermore, the combination of lithium and CL-A showed an

up-regulation of *cyclin-B1* mRNA while *cdc2* mRNA levels remained constant in both treated and untreated HL-60 cells.

To corroborate the RT-PCR data, we present evidence by Western blot analysis that Bcl-2 family of proteins and cell cycle regulatory genes indeed play a critical role in the regulation of apoptosis in HL-60 cells. Western blot analysis revealed a down-regulation of Bcl-2 under all treatment conditions. However, lithium and CL-A alone failed to show any detectable expression levels of both Bax and cyclin-B1 proteins. In contrast, the combination of both lithium and CL-A showed an up-regulation of Bax and Cdc2 proteins in HL-60 cells. These findings suggest that the molecular mechanism elicited by lithium, CL-A and the combination of both on the growth inhibition of HL-60 cells involves an aberrant expression of apoptotic and cell cycle regulatory genes. In addition, these observations may allude to a notion that both lithium and CL-A may be used and administered successfully as positive alternative anticancer drugs.